

Investigating the Symbiotic Community of Coral Species in Puerto Rico

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Introduction

- Coral reefs in Puerto Rico have experienced bleaching due to rising ocean temperatures caused by climate change that destroy their symbiotic relationship with the algae called zooxanthellae (Smith 1978).
- Two coral species found in Puerto Rico are *Montastraea cavernosa* (great star coral) and *Orbicella faveolata* (mountainous star coral).
- Corals are believed to be able to modify their symbiont communities when facing temperature stress by either switching algae with the surrounding environment or shuffling their pre-existing background algae (Adaptive Bleaching Hypothesis; Buddemeier and Fautin, 1993)
- Recent advances in research (e.g. quantitative PCR) now allow for more detailed observations into the dynamics of symbiont shuffling.



Cayo Enrique, and Cayo Mario, in La Parguera, Puerto Rico, where the samples were collected. (Google Earth Pro)

Objectives

- Do the two Caribbean coral species *Montastraea cavernosa* and *Orbicella faveolata* exhibit the same symbiont clades?
- Compare 2 species of corals' symbiotic algae communities (clades C and D), to gain a better understanding of the symbiotic community for coral conservation.
- Will both species exhibit clades C and D, as seen in previous research by Cunning and Baker 2013, who investigated the symbiotic algae community in Florida?



Image of *Orbicella faveolata*
Photo Credit: Florida Keys National Marine Sanctuary

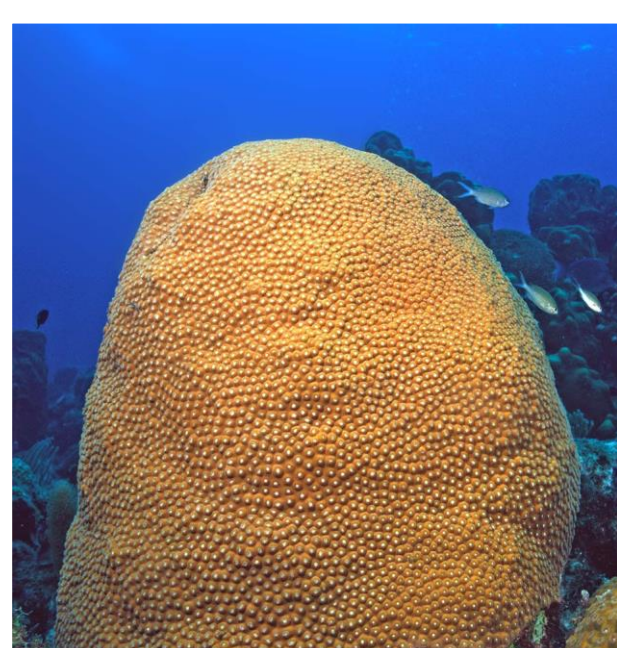


Image of *Montastraea cavernosa*
Photo Credit: Marion Haarsma

Methods



- DNA was extracted from twelve coral tissue samples of both coral species (sites 4, 5, and 6), following Cunning and Baker's modified organic extraction protocol (2013)
- Procedure involves incubating the samples at 1% SDS at 65°C, Proteinase K digest at 55°C, incubation with 1% CTAB at 65°C, mixing chloroform, 100% ethanol precipitation at -20°C, adding 0.3M sodium acetate to dried pellet and then second ethanol precipitation at -20°C, ethanol wash in 70% ethanol, and then Tris-EDTA buffer (TE) was added. Samples were stored at -20°C.

Methods (Continued)

- After this, the DNA is ready for quantitative polymerase chain reaction (qPCR) to identify the symbionts.
- The qPCR protocol we followed was adapted from Cunning and Baker 2013: all reactions were carried out in 10 µL volumes (with 5 µL Taqman Genotyping MasterMix and 2 µL DNA template) on a Roche LightCycler 96 Instrument.
- Thermal cycling conditions were set at: an initial incubation at 50°C for 2 min and 95°C for 10 min, followed by 40 cycles of 95°C for 10s and 60°C for 1 min.

Sample Name	1	2	3	4	5	6	7	8	9	10	11	12
4 OF 1	None	None	None	None	None	None	None	None	D	D	D	D
4 OF 2	None	None	None	None	None	None	None	None	D	D	D	D
4 OF 3	None	None	None	None	None	None	None	None	D	D	D	D
4 OF 4	None	None	None	None	None	None	None	None	D	D	D	D
4 OF 5	None	None	None	None	None	None	None	None	D	D	D	D
4 OF 6	None	None	None	None	None	None	None	None	D	D	D	D
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4 OF 12	None	None	None	None	None	None	None	None	D	D	D	D
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Fig. 1: Image is depicting a qPCR assay we set up for each coral species to determine if clade C or D was present.

Results

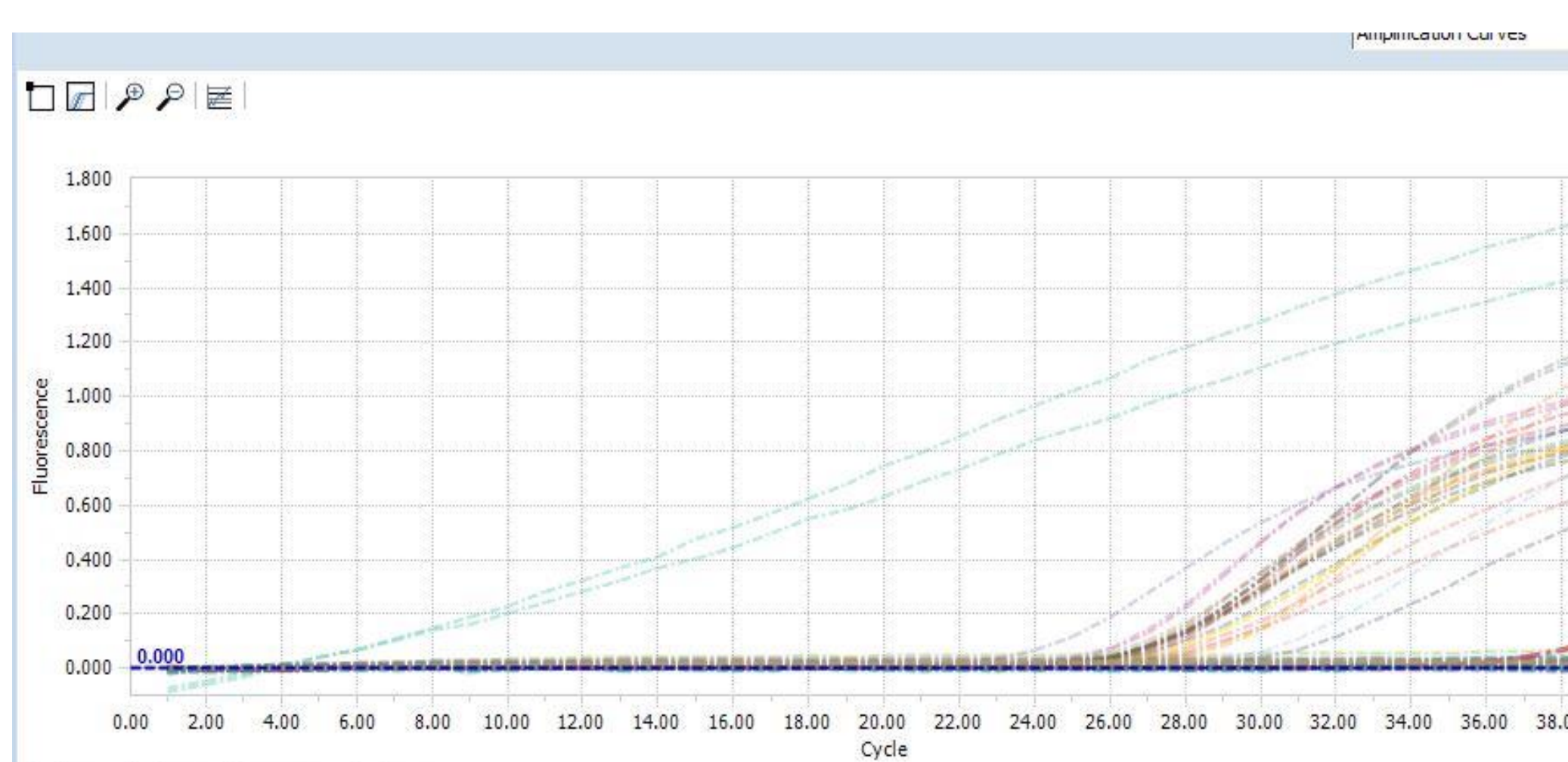


Fig. 2: Fluorescence graph representing the quantitative polymerase chain reaction process for the *M. cavernosa* samples. The curves indicate amplification.

Coral Species	Clade C				Clade D			
	1	2	3	4	1	2	3	4
Site 4	Control	Control	Control	Control	Control	Control	Control	Control
Site 5	Positive	Positive	Positive	Positive	Negative	Negative	Negative	Negative
Site 6	Positive	Positive	Positive	Positive	Negative	Negative		